

Green Corrosion Inhibitors for Mild Steel in H₂SO₄ Solutions (Period 2019-2022) - A Review

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Abstract

MS is the most commonly used alloy in industries, due to its remarkable features such as high thermal requirements, low cost, easy availability, high strength, durability and electrical conductivity. In recent years, scientists have focused on green inhibitors obtained from plants, fruit extracts and essential oils. Besides being environmentally friendly, in terms of corrosion resistance, plant extracts are becoming increasingly important, due to their low cost and toxicity, and high availability. Additionally, they are rich in organic compounds with polar atoms, such as O, P, S and N, containing multiple bonds in their molecules, through which they can adsorb onto the metal surface, forming a protective film, by various adsorption isotherms. This paper provides a review on research works done on MS corrosion control by naturally occurring plant extracts as corrosion inhibitors in H₂SO₄ solutions.

Keywords: corrosion inhibition; EIS; H₂SO₄; MS; plant extracts; PDP; WL.

Introduction*

Corrosion is the gradual destruction or deterioration of a metal by chemical or electrochemical interaction with an environment that leads to wastage of a metal surface. It occurs due to the metals spontaneous need to revert to a more stable form as it is found in nature. Corrosion is a continuous process and cannot be completely stopped. It causes financial implications in terms of product losses and replacement, and environmental pollution. Several techniques have been applied in order to reduce metallic corrosion. The use of inhibitors is one of the most practical and efficient methods for metals protection against corrosion [1].

The problems associated with traditional organic and inorganic inhibitors, which limit their usage nowadays, include biotoxicity, environmentally unfriendly properties, high cost and non-availability on demand. The use of natural products to inhibit corrosion is of enormous interest, since they are environmentally friendly, ecologically acceptable, readily available, inexpensive, and can be used nowadays as green inhibitors. Generally, plant extracts contain heteroatoms such as O, P, N and S. Adsorption of these atoms depends mainly on functional groups, steric factors, electron density at the donor atoms and also on the inhibitor's

*The abbreviations list is in page 175.

electronic structure. A corrosion inhibitor is generally referred to as a chemical substance that when applied in small quantities to a corrosive medium reduces the corrosion rate of a metal or an alloy [2]. Inhibitors retard metal corrosion by adsorbing onto a metallic surface. This process is influenced by some factors, which include the inhibitor molecular size and concentration, nature of substituents, test solution nature and T [3, 4]. The use of inhibitors for the control of metallic corrosion is one of the most practical approaches for preventing this phenomenon, especially in acid solutions, to prevent unexpected metal dissolution and acid consumption [4]. Due to inhibitor molecules adsorption onto metal surfaces, a protective film is formed, thus hindering corrosion. A great number of corrosion inhibitors have been studied for MS in acidic media.

MS contains 0.05 to 0.25 by weight of carbon, hence known as low CS. It is extensively applied in food, oil, chemical, energy and fabrication industries, due to its excellent mechanical properties. Therefore, MS is accorded the highest preference in all solutions to metals corrosion problems.

Acidic solutions are widely used in industry for removal of undesirable scale and rust in metal finishing. The most important areas of application are acid pickling, industrial acid cleaning and heat exchangers. H_2SO_4 is a strong acid, being used as a cleaner for rust, algae and scale from condensers and cooling towers [5].

Meanwhile, concerted efforts are being employed by scientists with a view to improve the life span of metallic and alloy materials, by finding suitable anti-corrosive compounds that can be used in various media to prevent metal dissolution.

Extraction methods

Various solvents have been used to obtain the desired concentration of active compounds from plant extracts. The efficient extraction of active compounds depends on the solvent used, among which the most common are water, ethanol, methanol, ethyl acetate, dichloromethane and hexane [6, 7]. Water could be the most convenient extraction solvent, since it is highly available, non-toxic, non-flammable and inexpensive [8].

T and immersion time

T has an important influence on the corrosion of metal surfaces. It is possible to modify the interaction between the corrosive medium and the metal surface in the presence of inhibitors. Some extracts exhibit an increasing IE(%) trend towards higher T [9]. However, other extracts have shown different behaviors. Thus, the evaluation of IE(%) as a function of T is important, since every extract could perform differently. Similarly, immersion time is another factor that could modify IE(%).

Characterization of techniques

A proper characterization of the extracts proposed as corrosion inhibitors is needed. Thus, several experimental techniques are available for this purpose. WL technique is based on the mass lost by corrosion, which is directly monitored to

assess the CR. Polarization tests, such as PDP, are based on the evaluation and analysis of the current produced by a variable potential in a working electrode [10]. EIS technique is used to determine the impedance of a system in terms of the frequency of a variable potential. EIS shows more information, for example, mechanisms and different resistances of the system. Surface characterization is commonly addressed by means of spectroscopy and microscopy techniques. SEM provides a clear comparison between the metal surface with and without a corrosion inhibitor, as well as other morphological information. Similarly, AFM obtains information regarding the shape of the metal surface for comparison purposes and topography imaging. Complementary characterizations are usually done through FT-IR, to obtain information on the functional groups and vibrational modes on corrosion inhibitors. Similarly, UV-vis helps to elucidate functional groups, electronic transitions and optical band gaps. XPS is a quantitative technique that is capable of identify elemental compositions and analyze the outermost molecular layers of a material. QCC has been recently used to analyze the adsorption mechanism of inhibitors on a metal surface.

Types of inhibitors

There are three types of inhibitors according to mechanism of electrochemical action. Anodic inhibitors slow down the oxidation reaction by blocking anodic sites (seat of metal oxidation), which decrease the density of the metal dissolution current and shift the corrosion potential in the positive direction. Cathodic corrosion inhibitors decrease the corrosion potential towards lower values, inhibiting the reactions that take place at the cathode, such as oxygen reduction and hydrogen evolution, shifting the corrosion potential in a negative direction. Mixed-type inhibitors act on both cathodic and anodic reactions.

Plant extracts as MS corrosion inhibitors

Table 1 shows recent studies, by various authors, on structural MS corrosion inhibition by plant extracts.

Table 1: Natural products as corrosion inhibitors for MS in a H₂SO₄ solution.

Plant	Corrosive medium	Techniques used	Type of inhibitor	IE(%)	Ref.
AC bark	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, AFM, UV-vis, FT-IR	Mixed	91.13 WL, 93.85 PDP, 93.12 EIS	[11]
AC bark	1 M H ₂ SO ₄	WL, OCP, PDP, SEM, UV, FT-IR	Mixed	93.96 WL, 98.91 PDP	[12]
AC pod	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, AFM, UV, QCC	Mixed	91.13 WL, 92.73 PDP, 94.98 EIS	[13]
AN	1 N H ₂ SO ₄	WL, PDP, FT-IR	--	--	[14]
AN leaves and stems	0.5 M H ₂ SO ₄	PDP, EIS, SEM, EDX, FT-IR, GC-MS	Mixed	AN leaves: 89.1 PDP, 88.8 EIS. AN stems: 80.0 PDP, 86.6 EIS	[15]
ASG	0.5 M H ₂ SO ₄ + 0.08 M KBr and KI	PDP, EIS	-	81.60 EIS with KI and 68.4 EIS with KBr	[16]
ACL	0.5 M H ₂ SO ₄	WL, PDP, EIS, FT-IR, SEM	Mixed	--	[17]
AE leaves	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, FT-IR, UV-Vis	---	96.0 WL	[18]
ANB	1 M H ₂ SO ₄	WL, OCP, PDP, FT-IR, UV-Vis	Mixed	71.94 WL, 90.00 PDP	[19]
AV leaves	0.5 M H ₂ SO ₄	WL, PDP, SEM	Mixed	88.9 WL	[20]

<i>Amaranthus</i>	4 N H ₂ SO ₄	WL, PDP, SEM	Cathodic	--	[21]
AS	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, AFM	Cathodic	--	[22]
AO	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, AFM, FT-IR	Mixed	91.62 PDP, 93.27 EIS	[23]
AH-AO	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, GC-MS	Mixed	85.0 WL, 88.0 PDP	[24]
AF-F leaves	1 M H ₂ SO ₄	WL, PDP, SEM, FT-IR	Mixed	85.49 WL, 88.39 PDP	[25]
PAM leaves	0.5 M H ₂ SO ₄	WL, SEM, EDX	--	97.89 WL	[26]
PAM seeds	0.75 M H ₂ SO ₄	WL, PDP, SEM, FT-IR	Mixed	74.55 WL, 68.37 PDP	[27]
AI leaves	1 M H ₂ SO ₄	WL	--	86.0 WL	[28]
<i>Berberine</i>	1M H ₂ SO ₄	WL, PDP, EIS, SEM, FT-IR, UV-Vis	Mixed	93.80 WL, 98.19 PDP, 98.02 EIS	[29]
BA	1M H ₂ SO ₄	WL, PDP, EIS, ATR-FTIR, SEM, EDX	Mixed	95.92 WL, 98.14 PDP	[30]
BK leaf	1.2 M H ₂ SO ₄	WL, FTIR, GC-MS, SEM, RSM, QCC	Mixed	93.0 WL, 87.6 PDP, 85.5 EIS	[31]
BM seeds	2 M H ₂ SO ₄	WL, HE, PDP, EIS, SEM, DFT, FT-IR, QCC	Mixed	93.48 WL, 89.27 PDP, 88.43 EIS, 94.83 HE	[32]
BE seeds	2.0 M H ₂ SO ₄	WL, SEM, FT-IR, DFT, QCC	--	96.0 WL	[33]
BE seeds	1.0 M H ₂ SO ₄	WL, SEM, FT-IR, QCC, DFT	--	95.53 WL	[34]
CN	2 M H ₂ SO ₄	WL, SEM, FT-IR, RSM	--	70.67 WL	[35]
CA leaves	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, AFM, FT-IR, UV-Vis	Mixed	93.81 WL, 96.46 PDP, 95.06 EIS	[36]
CB leaves	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM	Mixed	85.71 WL, 86.60 PDP, 86.86 EIS	[37]
CB leaves	1 M H ₂ SO ₄	WL	--	91.91 WL	[38]
CO stem	0.5 M H ₂ SO ₄	WL, SEM	--	94.34 WL	[39]
CM leaves	1 M H ₂ SO ₄	WL, PDP, EIS, SEM, FT-IR, EDX	Mixed	92.00 WL, 85.10 PDP	[40]
CNSA	1 M H ₂ SO ₄	WL, PDP, FT-IR, UV-Vis	Mixed	96.40 WL, 97.03 PDP	[41]
CS stem	1 M H ₂ SO ₄	WL, PDP	--	90.43 WL, 92.86 PDP	[42]
CP	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, UV-Vis, QMC	Mixed	98.09 WL, 84.09 PDP, 80.51 EIS	[43]
CuA leaf	1 M H ₂ SO ₄	WL, SEM, EDX, AFM, XRD, RSM	--	92.39 WL	[44]
CR	0.5 M H ₂ SO ₄	WL, PDP, EIS, GC-MS, FT-IR, AFM, DFT, QCC	Mixed	87.9 WL, 78.0 PDP, 77.0 EIS	[45]
DE	0.5 M H ₂ SO ₄	OCP, PDP, EIS, GC-MS, AFM, SEM, EDX	Mixed	68.47 PDP, 85.96 EIS	[46]
EO	3 M H ₂ SO ₄	WL, SEM, FT-IR, RSM	--	82.93 WL	[47]
EH	1 M H ₂ SO ₄	WL, PDP, EIS, FT-IR, SEM, EDX	Mixed	82.0 WL, 92.74 PDP, 86.28 EIS	[48]
<i>Eucalyptus</i> leaf	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, FT-IR	Mixed	--	[49]
EpO leaves	1 M H ₂ SO ₄	WL	--	92.72 WL	[50]
FH	1 M H ₂ SO ₄	WL, PDP, EIS, SEM, IR, UV-Vis	Mixed	86.56 WL, 85.14 PDP, 90.61 EIS	[51]
FPo and FPI	2 M H ₂ SO ₄	WL	--	FPO- 45.85 WL, FPL- 23.71 WL	[52]
GMM	0.5 M H ₂ SO ₄	WL, OCP, PDP, EIS, SEM, XPS, NMR, EPC, QCC	Mixed	95.4WL, 94.8 PDP, 95.4 EIS	[53]
Guava leaf	0.1, 0.3 and 0.5 M H ₂ SO ₄	WL, FT-IR	--	98.81 WL, 96.82 RSM	[54]
GEAH	1 M H ₂ SO ₄	WL, PDP, EIS, SEM, EDX, GC-MS, AFM and QCC	Mixed	78.57 WL, 95.89 PDP, 91.84 EIS	[55]
HBR leaves	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, EDX, FT-IR, ENA	Mixed	88.0 WL, 74.0 PDP, 82.0 EIS	[56]
<i>Hibiscus</i> leaves	0.5 M H ₂ SO ₄	WL, FT-IR	-	89.39 WL	[57]
HP	0.5 M H ₂ SO ₄	WL, FT-IR, SEM, GC-MS, UV-vis., MDS, QCC	--	95.11 WL	[58]
HSFS	0.5 M H ₂ SO ₄	PDP, EIS, SEM, FT-IR, UV-Vis	--	90.0 WL	[59]
HSPE and honey	1 M H ₂ SO ₄	PDP, GC-MS	Mixed	87.5 PDP	[60]

IC	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, ENA, FT-IR and UV-Vis, QCC	Mixed	77.96 WL, 87.02 PDP, 90.33 EIS	[61]
JC	1.0 M H ₂ SO ₄	WL, OCP, PDP	Mixed	92.05 WL, 91.34 PDP	[62]
JC leaves	4 M H ₂ SO ₄	WL	--	71.3 WL	[63]
JT stem bark	1 M H ₂ SO ₄	WL	--	75.61 WL	[64]
LD	5 M H ₂ SO ₄	WL, GM, FT-IR	--	82.63 WL, 84.09 GM	[65]
LH	0.5 M H ₂ SO ₄	WL, PDP	Mixed	--	[66]
LP	1 M H ₂ SO ₄	WL, PDP, EIS, SEM, FT-IR, AFM	Mixed	--	[67]
LC peels	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, UV-Vis, FT-IR, XRD	Mixed	97.8 WL, 95.7 PDP, 97.8 EIS	[68]
MK	1 M H ₂ SO ₄	WL, PDP, EIS, AAS, SEM, EDX, AFM, FT-IR, UV-Vis	Mixed	95.01 PDP	[69]
ME	0.4 and 2.5 M H ₂ SO ₄	WL, GM	--	62.07 WL	[70]
MH extract	0.5 M H ₂ SO ₄	WL, Colorometry, PDP, EIS, EDS, FESEM, FT-IR	Mixed	--	[71]
MAC	1 M H ₂ SO ₄	WL	--	90.0 WL	[72]
MP peels	H ₂ SO ₄	WL, PDP, EIS, FT-IR, UV-Vis, SEM, AFM, DFT	--	87.0 WL	[73]
MP stem extrude	1.0, 1.5 and 2.0 M H ₂ SO ₄	WL		84.75 WL	[74]
NI leaves	1.3 M H ₂ SO ₄	WL, PDP, EIS, SEM, FT-IR, QCC	Mixed	86.92 WL, 86.2 PDP, 86.0 EIS	[75]
NN leaves	1 M H ₂ SO ₄	WL, PDP, EIS	Mixed	76.47 WL, 75.56 PDP, 69.47 EIS	[76]
OEL leaves	1 M H ₂ SO ₄	PDP, EIS, SEM, FT-IR	Mixed	99.0 PDP, 96.0 EIS	[77]
OS leaves	1 N H ₂ SO ₄	WL, PDP, EIS, SEM, GC-MS, DFT, FT-IR, UV-Vis	Mixed	92.6 WL	[78]
Pichia sp. biofilm	1 M H ₂ SO ₄	WL, PDP, EIS, FT-IR, XRD, EDX, FE-SEM, XPS, AFM	Mixed	90.0 WL	[79]
PN leaves	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, RSM	Mixed	84.31 WL, 97.2 PDP, 85.34 EIS	[80]
PL bark	0.5 M H ₂ SO ₄	WL	--	88.0 WL	[81]
RLE	0.5 M H ₂ SO ₄	WL, PDP, EIS, SEM, QCC, FTIR, UV-Vis	Mixed	93.30 WL, 94.11 PDP, 92.19 EIS	[82]
RC plant	0.5 M H ₂ SO ₄	PDP, EIS, SEM, AFM, FTIR	Mixed	74.0 PDP, 75.0 EIS	[83]
RR	1 M H ₂ SO ₄	WL, PDP, SEM, EDX, FTIR	Mixed	87.51 WL, 93.24 PDP	[84]
Rice straw	1.5 M H ₂ SO ₄	WL, SEM	--	86.42 WL	[85]
SR leaf	1 N H ₂ SO ₄	WL, PDP, EIS, SEM, DFT, GC-MS, UV-Vis, FT-IR	Mixed	90.30 WL	[86]
SRB	1 M H ₂ SO ₄	WL, PDP, UV-Vis, FT-IR	Mixed	93.75 PDP	[87]
Siam weed	1 M H ₂ SO ₄	WL	--	95.0 WL	[88]
SX stem	1 M H ₂ SO ₄	WL, PDP, SEM, FT-IR	--	93.14 WL, 98.14 PDP	[89]
SC leaves	1 M H ₂ SO ₄ + KI	WL, GM, SEM	--	SC+ KI- 91.49 WL, 83.76 PDP	[90]
SA fruit	0.5 M H ₂ SO ₄	WL, PDP, EIS, AFM, SEM	---	93.25 WL	[91]
TT leaves	0.4, 0.5, 0.6 and 2.5 M H ₂ SO ₄	WL, GM	--	70.77 WL, 59.31 GM	[92]
TGL	0.5 M H ₂ SO ₄	WL, PDP, EIS, AFM, SEM	Mixed	96.70 WL	[93]
TP leaves	1 N H ₂ SO ₄	PDP, EIS, AFM, SEM, UV-Vis, FT-IR, NMR, GC-MS	Mixed	93.0 PDP	[94]
TC leaves	1 M H ₂ SO ₄	WL	--	64.0 WL	[95]
TI leaves	1 M H ₂ SO ₄	WL	--	92.47 WL	[96]
Thyme extract	0.5 M H ₂ SO ₄	WL, PDP, EIS	Mixed	98.0 WL	[97]
TCd	0.5 M H ₂ SO ₄	WL, PDP, RSM, BBD, EIS, SEM, AFM	Mixed	82.53 WL, 88.68 EIS	[98]
VA	1 M H ₂ SO ₄	WL, PDP, FT-IR, QCC	Mixed	89.11 WL	[99]
XA	5 M H ₂ SO ₄	WL, FT-IR, SEM	--	98.32 WL	[100]
ZM cobs	1 M H ₂ SO ₄	WL, PDP	Mixed	89.68 WL, 94.05 PDP	[101]
AV and ST	1 M H ₂ SO ₄	WL, PDP, SEM, FT-IR	Mixed	AV- 92.58, WL 88.06 PDP, ST- 90.79 WL, 83.22 PP	[102]
CT and JC	4 M H ₂ SO ₄	WL, GM	--	CT- 49.59 WL, 68.9 GM, JC- 55.77 WL, 61.30 GM	[103]
<i>Lavandula</i> and RCu oil	5 M H ₂ SO ₄	WL, PDP	--	96.35 WL, 90.00 PDP	[104]

AC: *Acacia catechu*; ACL: *Adina cordifolia* leaves; ACo: *Acacia concinna*; AE: *Ailanthus excelsa*; AF-F: *Athyrium filix-femina*; AH-AO: *Artemisia herba-alba* oil; AI: *Azadirachta indica*; AV: *Aloe vera*; AN: *Acacia nilotica*; ANB: *Alnus nepalensis* bark; AO: *Artabotrys odoratissimus*; AS: *Annona squamosa*; ASG: *Acacia senegal* gum, BA: *Berberis aristata*; BE: *Brachystegia eurycoma*; BK: *Bitter kola*; BM: *Black mustard*; CA: *Citrus aurantifolia*; CB: *Commelina benghalensis*; CM: *Cordia millenii*; CN: chicken nail; CNSA: *Coriaria nepalensis* stem alkaloid; CO: *Corchorus olitorius*; CP: *Croton persimilis*; CR: *Cyperus rotundus*; CS: *Crotalaria spectabilis*; CT: *Cascabela thevetia*; CuA: *Cucumis anguria*; DE: *Delonix elata*; EH: *Equisetum hyemale*; EO: *Epiphyllum oxypetalum*; EpO: *Euphorbia heterophylla*; FH: *Ficus hispida*; FPl: *Ficus platyphylla*; FPo: *Ficus polita*; GEAH: gum exudate from *Araucaria heterophylla*; GMM: *Glycine max* meal (Soybean); HBR: *Hardwickia binata* Roxb; HP: *Honeycomb propolis*; HSFS: *Hymenaea stigonocarpa* fruit shell; HSPE: *Hyptis suaveolens* poit extract; IC: *Ixora coccinea*; JC: *Jatropha curcas*; JT: *Jatropha tanjorensis*; LC: *Litchi chinensis*; LD: *Landolphia dulcis*; LH: *Landolphia heudelotii*; LP: *Liriope platyphylla*; MAC: *Musa acuminata* colla (native banana); ME: *Milicia excelsa*; MH: *Mitracarpus hirtus*; MK: *Magnolia kobus*; MP: *Musa paradisiaca*; NI: *Napoleonaea Imperialis*; PAM: *Persea americana* mill (avocado); NN: *Nelumbo nucifera*; OEL: *Olea europaea* L.; OS: *Oxalis stricta*; PL: *Polyalthia longifolia*; PN: *Picralima nitida*; RC: *Rhus coriaria*; RCu: *Ricinus communis*; RLE: Radish leaf extract; RR: *Rhynchosyris retusa*; SA: *Syzygium aromaticum*; SC: *Spondias cytheria*; SR: *Senegalia rugata*; SRB: *Shorea robusta* bark; SX: *Solanum xanthocarpum*; ST: *solanum tuberosum*; TC: *Terminalia catappa*; TCd: *Tinospora cordifolia*; TGL: *Tectona grandis* L; TI: *Terminalia ivorensis*; TP: *Tephrosia purpurea*; TT: *Talinum triangulare* (water leaf); VA: *Vernonia amygdalina* (bitter leaf); XA: *Xylopi aethiopica*; ZM: *Zea mays*.

Active phytoconstituents present in green inhibitors

In this review, most plant extracts were found to contain active phytochemicals such as tannins, flavonoids, saponins, alkaloids, phenols and glycosides. Some few plant extracts contain steroids, amino acids, carbohydrates, proteins, terpenes and β -sitosterol etc., which have a corrosion inhibitive effect.

Conclusions

This review paper has summarized the research works carried out by various researchers on MS corrosion in H_2SO_4 and on its inhibition by using different plant extracts and techniques. Many variables can be explored to evaluate a plant extract as corrosion inhibitor: concentration, extraction solvent, T and immersion time. In fact, plant extracts have numerous phytochemical constituents that are able to easily be adsorbed onto metals, thus inhibiting their CR by forming a passive film or an adsorbed layer that acted as a barrier. According to WL, PDP and EIS data, it is obvious that, with higher inhibitor concentration, CR decreases while IE(%) increases. An increase in T resulted in lower IE(%) of the tested products. Plants extracts can control metallic corrosion by inhibitory mechanisms (anodic, cathodic and mixed) in acidic environments. In this review, Langmuir's adsorption isotherm was found to be the most common. The effectiveness of the corrosion inhibitor was evaluated by WL, PDP and EIS. Other methods like SEM, FT-IR, UV-vis, RSM, AFM, GC-MS and EDX were also used to study the metals surfaces. Results obtained from WL data were in good agreement with PDP and EIS methods. This review may be useful to green corrosion chemists and researchers in the near future.

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Author's tasks

Conceived and designed the analysis; collected the data; provided data or analysis tools; performed the analysis; wrote the paper.

Abbreviations

AAS: atomic absorption spectroscopy

AFM: atomic force microscope

BBD: box-Behnken design

CR: corrosion rate

CS: carbon steel

DFT: density functional theory

EDS: energy dispersive spectroscopy

EDX: energy-dispersive X-ray spectroscopy

EIS: electrochemical impedance spectroscopy

ENA: electrochemical noise analysis

EPC: electro-phorosis characterization

FT-IR: fourier-transform infrared spectroscopy

GC-MS: gas chromatography mass spectrometry

GM: gasometry method

H₂SO₄: sulfuric acid

HE: hydrogen evolution

IE(%): inhibition efficiency

KBr: potassium bromide

KI: potassium iodide

MDS: molecular dynamics simulation

MS: mild steel

NaCl: sodium chloride

NaOH: sodium hydroxide

NMR: nuclear magnetic resonance

OCP: open circuit potential

PDP: potentiodynamic polarization

QCC: quantum chemical calculations

QMC: quantum mechanical calculations

RS: Raman spectroscopy

RSM: response surface methodology

SE: synergistic effect

SEM: scanning electron microscopy

T: temperature

UV-vis: ultraviolet-visible spectroscopy or ultraviolet-visible spectrophotometry

WL: weight loss

XPS: X-ray photoelectron spectroscopy

XRD: X-ray diffraction

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